dicom_csv

NeuroML

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This is a collection of utils for gathering, aggregation and handling metadata from DICOM files.

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1.1 Crawling

Contains functions for gathering metadata from individual DICOM files or entire directories.

```
dicom_csv.crawler.get_file_meta(path: Union[Path, str], force: bool = True, read\_pixel\_array: bool = False, unpack\_volumetric: bool = False, extract\_private: bool = False) \rightarrow Iterable[dict]
```

Get a dict containing the metadata from the DICOM file located at path.

Parameters

```
PathLike (path -) – full path to file
```

:param : full path to file :param force - bool: pydicom.filereader.dcmread force parameter, default is False :param : pydicom.filereader.dcmread force parameter, default is False :param read_pixel_array - bool: if True, crawler will add information about DICOM pixel_array, False significantly increases crawling time,

default is True.

:param

[if True, crawler will add information about DICOM pixel_array, False significantly increases crawling time,] default is True.

Notes

The following keys are added:

NoError: whether an exception was raised during reading the file.

HasPixelArray: (if NoError is True) whether the file contains a pixel array. PixelArrayShape: (if HasPixelArray is True) the shape of the pixel array.

For some formats the following packages might be required:

```
>>> conda install -c glueviz gdcm # Python 3.5 and 3.6
>>> conda install -c conda-forge gdcm # Python 3.7
```

```
dicom_csv.crawler.join_tree(top: Union[Path, str], ignore_extensions: Sequence[str] = (), relative: bool = True, verbose: int = 0, read_pixel_array: bool = False, force: bool = True, unpack_volumetric: bool = True, extract_private: bool = False, total: bool = False) \rightarrow DataFrame
```

Returns a dataframe containing metadata for each file in all the subfolders of top.

Parameters

- top (PathLike) path to crawled folder
- ignore_extensions (Sequence) list of extensions to skip during crawling
- **relative** (*bool*) whether the PathToFolder attribute should be relative to top default is True.
- verbose (int) -

the verbosity level:

- 0 no progressbar
- 1 progressbar with iterations count
- 2 progressbar with filenames
- **total** (*bool*) whether to show the total number of files in the progressbar. This is adds a bit of overhead, because each file will be visited a second time (without being opened).

References

See the Working with DICOM files tutorial for more details.

Notes

The following columns are added:

NoError: whether an exception was raised during reading the file.

HasPixelArray:(if NoError is True) whether the file contains a pixel array(added if read_pixel_array is True).

PixelArrayShape: (if HasPixelArray is True) the shape of the pixel array (added if read_pixel_array is True).

PathToFolder

FileName

For some formats the following packages might be required:

```
>>> conda install -c glueviz gdcm # Python 3.5 and 3.6
>>> conda install -c conda-forge gdcm # Python 3.7
```

1.2 Aggregation

Tools for grouping DICOM metadata into images.

```
\label{local_com_csv} \begin{subarrate}{ll} dicom\_csv.aggregate\_images(\textit{metadata: DataFrame, by: Union[str, Sequence[str]], } \\ process\_series: Optional[Callable] = None) \to DataFrame \end{subarrate}
```

Groups DICOM metadata into images (series).

Parameters

• **metadata** – a dataframe with metadata returned by *join_tree*.

- by a list of column names by which the grouping will be performed. Default columns are: PatientID, SeriesInstanceUID, StudyInstanceUID, PathToFolder, PixelArrayShape, SequenceName
- process_series a function that processes an aggregated series before it will be joined into a single entry

References

See the Working with DICOM files tutorial for more details.

Notes

The following columns are added:

SlicesCount: the number of files/slices in the image. FileNames: a list of slash ("/") separated file names.

InstanceNumbers: (if InstanceNumber is in columns) a list of comma separated InstanceNumber values.

The following columns are removed:

FileName (replaced by FileNames), InstanceNumber (replaced by InstanceNumbers), any other columns that differ from file to file.

 $dicom_csv.aggregation.normalize_identifiers(metadata: DataFrame) \rightarrow DataFrame$ Converts PatientID to str and fills nan values in SequenceName.

Notes

The input dataframe will be mutated.

 $dicom_csv.aggregation.select(dataframe: DataFrame, query: str, **where: str) \rightarrow DataFrame$

1.3 Loading

```
dicom_csv.misc.get_image(instance: Dataset, to_color_space: Optional[str] = None)
dicom_csv.misc.stack_images(series: Sequence[Dataset], axis: int = -1, to_color_space: Optional[str] = None)
```

1.4 Spatial operations

 $dicom_csv.spatial.get_orientation_matrix(series: Sequence[Dataset]) \rightarrow ndarray$

Returns a 3 x 3 orthogonal transition matrix from the image-based basis to the patient-based basis. Rows are coordinates of image-based basis vectors in the patient-based basis. Columns are coordinates of patient-based basis vectors in the image-based basis vectors.

See https://dicom.innolitics.com/ciods/rt-dose/image-plane/00200037 for details.

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```
dicom_csv.spatial.get_slice_plane(instance: Dataset) \rightarrow Plane
dicom\_csv.spatial.get\_slices\_plane(series: Sequence[Dataset]) \rightarrow Plane
class dicom_csv.spatial.Plane(value)
     Bases: Enum
     An enumeration.
     Sagittal = 0
     Coronal = 1
     Axial = 2
dicom_csv.spatial.order_series(series: Sequence[Dataset], decreasing: bool = True) \rightarrow Sequence[Dataset]
dicom_csv.spatial.get_slice_locations(series: Sequence[Dataset]) \rightarrow ndarray
     Computes slices location from ImagePositionPatient. NOTE: the order of slice locations can be both increasing
     or decreasing for ordered series (see order series).
dicom_csv.spatial.locations_to_spacing(locations: Sequence[float], max_delta: float = 0.1, errors: bool =
                                                True) \rightarrow float
dicom_csv.spatial.get_slice_spacing(series: Sequence[Dataset], max_delta: float = 0.1, errors: bool =
                                            True) \rightarrow float
     Returns constant distance between slices of a series. If the series doesn't have constant spacing - raises ValueError
     if errors is True, returns np.nan otherwise.
dicom_csv.spatial.get_pixel_spacing(series: Sequence[Dataset]) → Tuple[float, float]
     Returns pixel spacing (two numbers) in mm.
dicom_csv.spatial.get_voxel_spacing(series: Sequence[Dataset]) \rightarrow Tuple[float, float]
     Returns voxel spacing: pixel spacing and distance between slices' centers.
dicom_csv.spatial.get_image_position_patient(series: Sequence[Dataset]) → ndarray
     Returns ImagePositionPatient stacked into array.
dicom_csv.spatial.drop_duplicated_slices(series: Sequence[Dataset], tolerance_hu=1) \rightarrow
                                                  Sequence[Dataset]
dicom\_csv.spatial.orientation\_matrix\_to\_slices\_plane(om: ndarray) \rightarrow Plane
dicom_csv.spatial.get_slice_orientation(*args, **kwds)
      get_slice_orientation is deprecated!
dicom\_csv.spatial.get\_slices\_orientation(series: Sequence[Dataset]) \rightarrow SlicesOrientation
      get_slices_orientation is deprecated!
class dicom_csv.spatial.SlicesOrientation(transpose: bool, flip_axes: tuple)
     Bases: tuple
     Defines how slices should be transformed in order to be canonically oriented: First transpose slices if transpose
     == True. Then flip slices along flip_axes (they already account for transposition).
     property transpose
           Alias for field number 0
```

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property flip_axes

Alias for field number 1

```
dicom_csv.spatial.orientation_matrix_to_slices_orientation(*args, **kwds)
```

orientation_matrix_to_slices_orientation is deprecated!

```
dicom_csv.spatial.get_axes_permutation(*args, **kwds)
```

get_axes_permutation is deprecated!

dicom_csv.spatial.get_flipped_axes(*args, **kwargs)

<lambda> is deprecated!

 $dicom_csv.spatial.get_image_plane(series: Sequence[Dataset]) \rightarrow Plane$

get_image_plane is deprecated!

dicom_csv.spatial.restore_orientation_matrix(metadata: Union[Series, DataFrame])

Fills nan values (if possible) in metadata's ImageOrientationPatient* rows.

Required columns: ImageOrientationPatient[0-5]

Notes

The input dataframe will be mutated.

1.5 Console scripts

This library contains a console script around <code>join_tree</code>, which is added to your namespace after installation:

```
dicom-csv folder/with/dicoms path/to/metadata.csv

# pass --help for more details:
dicom-csv --help
```

1.6 Working with DICOM files

Before we start analysing our files, let's install some additional libraries, which add support for various medical imaging formats:

```
conda install -c glueviz gdcm # Python 3.5 and 3.6 conda install -c conda-forge gdcm # Python 3.7
```

We'll be working with a subset of the CT Lymph Nodes dataset which can be downloaded here.

```
path = '~/dicom_data/'
```

1.6.1 Crawling

join_tree is the main function that collects the DICOM files' metadata:

```
from dicom_csv import join_tree

df = join_tree(path, relative=True, verbose=False)
```

It recursively visits the subfolders of path, also it adds some additional attributes: NoError, HasPixelArray, PathToFolder, FileName:

```
len(df), df.NoError.sum(), df.HasPixelArray.sum()
```

```
(2588, 2587, 2587)
```

Thre resulting dataframe has 2588 files' metadata in it, and only one file was openned with errors, let's check which one:

```
df.loc[~df.NoError, ['FileName', 'PathToFolder']]
```

There is a file readme.txt in the root of the folders tree, which is obvisously not a DICOM file.

Note that PathToFolder is relative to path, this is because we passed relative=True to join_tree.

```
# leave only dicoms that contain images (Pixel Arrays)
dicoms = df[df.NoError & df.HasPixelArray]
dicoms.FileName[1], dicoms.PathToFolder[1]
```

```
('000466.dcm',
'ABD_LYMPH_001/09-14-2014-ABDLYMPH001-abdominallymphnodes-30274/abdominallymphnodes-
→26828')
```

1.6.2 Aggregation

Next, we can join the dicom files into series, which are often easier to operate with:

```
from dicom_csv import aggregate_images
images = aggregate_images(dicoms)
len(images)
```

```
4
```

aggregate_images also adds some attributes: SlicesCount, FileNames, InstanceNumbers, check its docstring for more information.

For example FileNames contains all the files that are part of a particular series:

```
images.FileNames[0][:50] + '...'
```

```
'000466.dcm/000312.dcm/000150.dcm/000357.dcm/000311...'
```

As you can see, they are not ordered by default, but you can change this behaviour by passing the process_series argument which receives a subset of the dataframe, containing files from the same series:

```
'000000.dcm/000001.dcm/000002.dcm/000003.dcm/000004...'
```

1.6.3 Loading

You can load a particular series' images stacked into a numpy array using the following function:

```
img = load_series(images.loc[0], path)
```

it expects a row from the aggregated dataframe and, optionally, the path argument, if the paths are relative.

The image's orientation as well as the slices' order can be determined automatically, if you pass orientation=True:

```
img = load_series(images.loc[0], path, orientation=True)
```

```
print(img.shape, images.PixelArrayShape[0], images.SlicesCount[0])
```

```
(512, 512, 661) 512,512 661
```

1.7 Loading contours stored in DICOM format (RTstructure).

Segmentation mask is stored in DICOMs in as a set of plain contours. These contours are nothing but a 2D curves defined by set of coordinates in 3D space. Specifically these coordinates are in physical space not in voxel space. Every single RTStructure might contain contours for multiple masks (e.g. Brain, Left_Eye, GTV etc.).

dicom-csv contains several functions to collect contours stored in RTstructures and move them into image's voxel space.

1.7.1 Crawling images folder

join_tree is the main function that collects the DICOM files' metadata:

```
from dicom_csv import join_tree

df = join_tree(path, relative=False, verbose=False)
```

1.7.2 Select rows related to RTStructures

collect_rtstruct is the function which selects rows related to RTstructures and copies additional metadata from corresponding images (RTstructure itself does not contain information about spatial orientation, it only contains coordinates and link to corresponding DICOM image):

```
from dicom_csv.rtstructs.csv import collect_rtstructs

df_rtstructs = collect_rtstructs(df)
```

1.7.3 Extract contours coordinates from RTStructure

contours_dict extract all coordinates from a single row of df_rtstructs:

```
from dicom_csv.rtstruct.contour import read_rtstruct
contours_dict = read_rtstruct(df_rtstructs.iloc[0])
```

It outputs python dictionary with keys - names of the contours and values - $n \times 3$ nd.arrays of coordinates in physical space.

1.7.4 Move contours to voxel space

Finally, contours_image_dict moves these coordinates into voxel space (basis change transformation, essentially rotation, translation and stretching):

```
from dicom_csv.rtstruct.contour import contours_image_dict
contours_image_dict = contours_to_image(df_rtstructs.iloc[0], contours_dict)
```

Resulting dictionary contains all contours stored in corresponding RTStructure.

1.7.5 Contours to masks

TODO

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